

# REPORT DOCUMENTATION PAGE

Form Approved  
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of

1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE	3. REPORT TYPE AND DATES COVERED	
	March 1, 2002	Final Report	15 May 1998–31 Dec. 2000
4. TITLE AND SUBTITLE		5. FUNDING NUMBERS	
<b>Performance-Enhancing Biomolecular Treatment Strategies for Naval Graywater Filtration Systems</b>		N00014-98-1-0696	
6. AUTHOR(S)			
Douglas S. Clark			
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)		8. PERFORMING ORGANIZATION REPORT NUMBER	
Department of Chemical Engineering 201 Gilman Hall University of California Berkeley, CA 94720			
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)		10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
Office of Naval Research 800 N. Quincy St. Arlington, VA 22217-5000			
11. SUPPLEMENTARY NOTES			
12a. DISTRIBUTION / AVAILABILITY STATEMENT			
Distribution Unlimited			
20020325 177			
13. ABSTRACT (Maximum 200 words)			
<p>The primary objective of this research was to examine the feasibility of using immobilized-enzyme membranes to improve the performance and lifetime of a graywater filtration system. Enzyme-treated membranes were examined for increased flux relative to membranes without enzyme treatment. Of fifteen hydrolytic enzymes (seven proteases, six lipases, and two amylases) screened in soluble form for the ability to degrade synthetic graywater, Protease X from <i>Bacillus thermoproteolyticus rokko</i> showed the highest activity, and several enzymes (e.g., lipase Type II from porcine pancreas, and <math>\alpha</math>-chymotrypsin from bovine pancreas) showed significant but lower activity. We also developed a new methodology for preparing biocatalytic films and paints based on polydimethylsiloxane (PDMS), which might be used as anti-fouling treatments for a wide variety of materials, including filters and membranes. Such enzyme treatments may increase the service lifetime and efficiency of filter membranes, resulting in dramatic cost savings and minimal maintenance for an eventual on-board graywater treatment system.</p>			
14. SUBJECT TERMS		15. NUMBER OF PAGES	
Graywater, filtration, immobilized enzyme membranes		4	
		16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT	18. SECURITY CLASSIFICATION OF THIS PAGE	19. SECURITY CLASSIFICATION OF ABSTRACT	20. LIMITATION OF ABSTRACT
Unclassified	Unclassified	Unclassified	UL

## GENERAL INSTRUCTIONS FOR COMPLETING SF 298

The Report Documentation Page (RDP) is used in announcing and cataloging reports. It is important that this information be consistent with the rest of the report, particularly the cover and title page. Instructions for filling in each block of the form follow. It is important to *stay within the lines* to meet *optical scanning requirements*.

**Block 1. Agency Use Only (Leave blank).**

**Block 2. Report Date.** Full publication date including day, month, and year, if available (e.g. 1 Jan 88). Must cite at least the year.

**Block 3. Type of Report and Dates Covered.** State whether report is interim, final, etc. If applicable, enter inclusive report dates (e.g. 10 June 87 - 30 June 88).

**Block 4. Title and Subtitle.** A title is taken from the part of the report that provides the most meaningful and complete information. When a report is prepared in more than one volume, repeat the primary title, add volume number, and include subtitle for the specific volume. On classified documents enter the title classification in parentheses.

**Block 5. Funding Numbers.** To include contract and grant numbers; may include program element number(s), project number(s), task number(s), and work unit number(s). Use the following labels:

C - Contract  
G - Grant  
PE - Program Element

PR - Project  
TA - Task  
WU - Work Unit  
Accession No.

**Block 6. Author(s).** Name(s) of person(s) responsible for writing the report, performing the research, or credited with the content of the report. If editor or compiler, this should follow the name(s).

**Block 7. Performing Organization Name(s) and Address(es).** Self-explanatory.

**Block 8. Performing Organization Report Number.** Enter the unique alphanumeric report number(s) assigned by the organization performing the report.

**Block 9. Sponsoring/Monitoring Agency Name(s) and Address(es).** Self-explanatory.

**Block 10. Sponsoring/Monitoring Agency Report Number.** *(If known)*

**Block 11. Supplementary Notes.** Enter information not included elsewhere such as: Prepared in cooperation with...; Trans. of ...; To be published... When a report is revised, include a statement whether the new report supersedes or supplements the older report.

**Block 12a. Distribution/Availability Statement.** Denotes public availability or limitations. Cite any availability to the public. Enter additional limitations or special markings in all capitals (e.g. NOFORN, REL, ITAR).

DOD - See DODD 5230.24, "Distribution Statements on Technical Documents."

DOE - See authorities.

NASA - See Handbook NHB 2200.2.

NTIS - Leave blank.

**Block 12b. Distribution Code.**

DOD - Leave blank.

DOE - Enter DOE distribution categories from the Standard Distribution for Unclassified Scientific and Technical Reports.

NASA - Leave blank.

NTIS - Leave blank.

**Block 13. Abstract.** Include a brief (*Maximum 200 words*) factual summary of the most significant information contained in the report.

**Block 14. Subject Terms.** Keywords or phrases identifying major subjects in the report.

**Block 15. Number of Pages.** Enter the total number of pages.

**Block 16. Price Code.** Enter appropriate price code (*NT/S only*).

**Blocks 17. - 19. Security Classification.** Self-explanatory. Enter U. S. Security Classification in accordance with the U.S. Security Regulations (i.e., UNCLASSIFIED). If form contains classified information, stamp classification on the top and bottom of the page.

**Block 20. Limitation of Abstract.** This block must be completed to assign a limitation to the abstract. Enter either UL (unlimited) or SAR (same as report). An entry in this block is necessary if the abstract is to be limited. If blank, the abstract is assumed to be unlimited.

FINAL REPORT

GRANT #: N00014-98-1-0696

PRINCIPAL INVESTIGATOR: Dr. Douglas S. Clark

INSTITUTION: University of California, Berkeley

GRANT TITLE: Performance-Enhancing Biomolecular Treatment Strategies for Naval Graywater Filtration Systems

AWARD PERIOD: 15 May 1998 - 31 December 2000

OBJECTIVE: To examine the feasibility of using immobilized-enzyme membranes to improve the performance and lifetime of a graywater filtration system.

APPROACH: Enzyme-treated membranes were examined for increased flux relative to membranes without enzyme treatment. As part of a three-way research project, two strategies were pursued at U.C. Berkeley: enzyme immobilization on the filter membrane surface via direct adsorption or covalent attachment, and enzyme immobilization via surface treatments with siloxane films. For example, polydimethylsiloxane, which is known to resist biological fouling, was used in the preparation of immobilized-enzyme silicone coatings for membrane-surface treatment. Once appropriate enzymes were identified, they were immobilized for testing against synthetic graywater.

ACCOMPLISHMENTS: We have designed and constructed a 96-channel filtration system for high throughput flux measurements with immobilized-enzyme membranes, enabling up to 96 enzymes and/or their combinations and immobilization methods to be screened simultaneously. Different enzymes and their combinations were screened for their ability to inhibit membrane fouling by graywater.

Of fifteen hydrolytic enzymes (seven proteases, six lipases, and two amylases) screened in soluble form for the ability to degrade synthetic graywater, Protease X from *Bacillus thermoproteolyticus* rokko showed the highest activity, and several enzymes (e.g., lipase Type II from porcine pancreas, and  $\alpha$ -chymotrypsin from bovine pancreas) showed significant but lower activity.  $\alpha$ -Chymotrypsin was then immobilized to PVDF ultrafiltration membranes

(molecular weight cutoff: 10,000) by seven different methods, with physical adsorption yielding the highest activity of immobilized enzyme for the hydrolysis of the model substrate benzoyl L-tyrosine ethyl ester. Based on this result, the enzymes were immobilized by physical adsorption to PVDF membranes, and the immobilized-enzyme membranes evaluated for flux improvement in filtration studies with synthetic graywater. As shown in Table I below, Protease X again had the greatest effect, producing a 70% improvement in flux over an 8-hour period.

**Table I. Improvement of permeate fluxes by immobilized enzymes**

Enzyme	Control	BSA	Trypsin	Chymotrypsin
Flux Gain (%)	0.0	0.6	9.3	38
Enzyme	Amylase <i>Bacillus</i> sp.	Amylase <i>B. licheniformis</i>	Lipase II (porcine)	Lipase AY30
Flux Gain (%)	0.7	6.5	49	16
Enzyme	Capalase (Lipase)	Lipase M	Lipase E.D.	Protease XXIII <i>A. oryzae</i>
Flux Gain (%)	6.9	14	43	23
Enzyme	Protease XIV <i>S. griseus</i>	Subtilisin Carlsberg	Italase (Lipase)	Protease K <i>T. album</i>
Flux Gain (%)	23	24	16	18
Enzyme	Protease X			
Flux Gain (%)	70			

In addition, a new type of ultrafiltration membrane, prepared from blended poly(vinylidene fluoride) (PVDF) and poly(MMA-co-MA) containing protease X (from *Bacillus thermoproteolyticus rokko*) was screened for antifouling activity against graywater. We also developed a new methodology for preparing biocatalytic films and paints

based on polydimethylsiloxane (PDMS), which might be used as anti-fouling treatments for a wide variety of materials, including filters and membranes.

To this end, the hydrolytic enzymes pronase and  $\alpha$ -chymotrypsin were immobilized by either sol-gel entrapment or by a covalent attachment method into a polydimethylsiloxane (PDMS) matrix and cast into thin films or into an oil-based paint formulation. All of the coatings retained enzymatic activity and adhered to several different materials. PDMS immobilized enzyme also exhibited higher thermostability than enzyme in solution or covalently attached to the outer surface of the PDMS. A porous membrane based on PDMS-immobilized enzyme was also prepared by an immersion precipitation process. Protein adsorption measurements showed that the enzyme containing films and paints adsorbed less protein than enzyme-free controls, and that protein adsorption decreased with increasing proteolytic activity of the coating. These coatings thus provide the means to apply a stable enzymatic surface to a wide range of materials, and may be generally useful as biocatalytic paints with antifouling properties.

Finally, this research has also laid the foundation for follow-up research to develop a practical filtration system for shipboard application. This follow-up project is a tripartite collaboration between the polymer membrane synthesis and characterization laboratory of Professor Benny Freeman at N.C. State, the enzyme technology laboratory of Professor Douglas S. Clark at U.C. Berkeley, and the thin-film composite membrane development facility at Membrane Technology and Research, Inc. Furthermore, the biocatalytic methodology developed in this project could conceivably be extended to antifouling paints, coatings, and films for use on ship hulls, in implantable materials, and as protective coatings for medical devices.

SIGNIFICANCE: Effective methods for the treatment of shipboard wastewater are of considerable importance as the Navy moves forward in the 21st Century. The enzyme immobilization methods developed thus far are an important first step toward reducing filter fouling and increasing the operational lifetime of the filter membranes. Such enzyme treatments may increase the service lifetime and efficiency of filter membranes, resulting in dramatic cost savings and minimal maintenance for an eventual on-board graywater treatment system. Moreover, the biocatalytic

method we propose offers the Navy an environmentally friendly way of accomplishing this task. The ultimate products of this research will be biocatalytic filtration systems that resist fouling during the shipboard filtration of graywater. Such devices may generate interest for commercialization, and may be appropriate as a new technology for development by a start-up venture.

#### PUBLICATIONS AND PRESENTATIONS

1. Y.-D. Kim, J. S. Dordick, and D. S. Clark (2001), "Siloxane-Based Biocatalytic Films and Paints: New Enzyme-Containing Coatings with Foulant-Resistant Properties," *Biotechnol. Bioeng.*, **72**, 475-482.
2. Y.-D. Kim, J. S. Dordick, and D. S. Clark, "Biocatalytic Polydimethylsiloxane Coatings," poster presented at Gordon Conference on Polymers (East), New London, CT, June, 2000.
3. Guo, Y. Yang, Y. D. Kim, and D. S. Clark, "High Throughput Screening of Hydrolytic Enzymes for Graywater Treatment and Prevention of Fouling," poster presented at ACS National Meeting, San Francisco, CA, March, 2000.